Electroacupuncture improves gut barrier dysfunction in prolonged hemorrhagic shock rats through vagus anti-inflammatory mechanism

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Abstract

AIM: To investigate whether electroacupuncture (EA) at Zusanli (ST36) prevents intestinal barrier and remote organ dysfunction following prolonged hemorrhagic shock through a vagus anti-inflammatory mechanism.

METHODS: Sprague-Dawley rats were subjected to about 45% of total blood volume loss followed by delayed fluid replacement (DFR) with Ringer lactate 3h after hemorrhage. In a first study, rats were randomly divided into six groups: (1) EAN: EA at non-channel acupoints followed by DFR; (2) EA: EA at ST36 after hemorrhage followed by DFR; (3) VGX/EA: vagotomy (VGX) before EA at ST36 and DFR; (4) VGX/EAN: VGX before EAN and DFR; (5) α-bungarotoxin (α-BGT)/EA: intraperitoneal injection of α-BGT before hemorrhage, followed by EA at ST36 and DFR; and (6) α-BGT/EAN: α-BGT injection before hemorrhage followed by EAN and DFR. Survival and mean arterial pressure (MAP) were monitored over the next 12 h. In a second study, with the same grouping and treatment, cytokine levels in plasma and intestine, organ parameters, gut injury score, gut permeability to 4 kDa FITC-dextran, and expression and distribution of tight junction protein ZO-1 were evaluated.

RESULTS: MAP was significantly lowered after blood loss; EA at ST36 improved the blood pressure at corresponding time points 3 and 12 h after hemorrhage. EA at ST36 reduced tumor necrosis factor-α and interleukin (IL)-6 levels in both plasma and intestine homogenates after blood loss and DFR, while vagotomy or intraperitoneal injection of α-BGT before EA at ST36 reversed its anti-inflammatory effects, and EA at ST36 did not influence IL-10 levels in plasma and intestine. EA at ST36 alleviated the injury of intestinal villus, the gut injury score being significantly lower than that of EAN group (1.85 ± 0.33 vs 3.78 ± 0.59, P < 0.05). EA at ST36 decreased intestinal permeability to FITC-dextran compared with EAN group (856.95 ng/mL ± 90.65 ng/mL vs 2305.62 ng/mL ± 278.32 ng/mL, P < 0.05). EA at ST36 significantly preserved ZO-1 protein expression and localization at 12 h after hemorrhage. However, EA at non-channel acupoints had no such effect, and abdominal vagotomy and α-BGT treatment could weaken or eliminate the effects of EA at ST36. Besides, EA at ST36 decreased blood aminotransferase, MB isoenzyme of creatine kinase and creatinine vs EAN group at corresponding time points. At the end of 12-h experiment, the survival rate of the EA group was significantly higher than that of the other groups.

CONCLUSION: EA at ST36 attenuates the systemic inflammatory response, protects intestinal barrier integrity, improves organ function and survival rate after prolonged hemorrhagic shock.
hemorrhagic shock via activating the cholinergic anti-inflammatory mechanism.

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Key words: Hemorrhagic shock; Zusanli; Electro-acupuncture; Intestinal permeability; Tight junction

Core tip: The most important novel findings from this study are that when delayed resuscitation is inevitable during emergency situations such as hemorrhagic shock occurring in war without sufficient fluids, electroacupuncture at ST36 can be performed and it can successfully attenuate systemic inflammation, decrease gut injury and permeability and improve blood pressure and outcomes, which is consistent with preserved intestinal barrier function after hemorrhage and delayed fluid resuscitation.


INTRODUCTION

The current treatment for hypovolemic shock focuses on maintaining sufficient tissue perfusion and vital organ function with early and adequate fluid replenishment. Delayed fluid resuscitation for hemorrhagic shock usually occurs when mass casualties happen in austere environments such as battlefield, earthquake, or accidents, where intravenous fluid resuscitation is often difficult or even impossible. Subsequent to delayed resuscitation of hypovolemic shock, a high mortality and an increase in incidence of serious complications such as systemic inflammatory response syndrome (SIRS) and the multiple organ dysfunction syndrome (MODS) may thus befall to these victims[1]. Although delayed resuscitation have been demonstrated to result in a more profound shock insult than early resuscitation[2,3], its pathological mechanisms remain poorly understood. One potential pathogenic mechanism appears to be associated with proinflammatory cytokine response[4,5] and, gut plays a key role in the development of intestinal and systemic inflammatory response following hemorrhagic shock and severe burn[6]. Gut becomes a source of proinflammatory mediators resulting from impairment of intestinal mucosal barrier that may amplify SIRS, produce a systemic response state and distant organ failure, and lead to MODS or even death[7-9]. Thus, interventions to prevent intestinal barrier breakdown, excessive inflammation and subsequent organ dysfunction when hemorrhagic shock prolonged due to delayed resuscitation are crucial in controlling hemorrhagic shock without sufficient fluid infusion, especially during evacuation and transportation.

The cholinergic anti-inflammatory pathway is a neural mechanism that inhibits the expression of pro-inflammatory cytokines through the interaction of the principle vagus nerve neurotransmitter, acetylcholine, and the cholinergic α7 nicotinic acetylcholine receptor (α7nAChR) subunit located on cytokine-expressing cells by stimulating the vagus nerve by either electrical or pharmacological methods[10,11]. Activation of the cholinergic anti-inflammatory pathway by vagus nerve stimulation can prevent cytokine release and tissue injury[12], prolong survival and protect against the development of hypotension in rats during lethal hemorrhagic shock[13]. Recently, researchers have demonstrated an expanded role for vagus nerve stimulation and the cholinergic anti-inflammatory mechanism that provides a protective effect on the gut against epithelial barrier dysfunction and alleviates inflammatory injury in intestine and remote organs[14-16]. However, due to complicated manipulation and untoward side effects, including serious tissue injury, it is still difficult to apply electrical stimulation to the vagus nerve in clinical practice. Therefore, a more clinically desirable alternative therapy needs to be established during the resuscitative phase of trauma care.

Acupuncture as one of the therapeutic maneuvers in traditional Chinese medicine (TCM) has been applied in clinics for thousands of years, and it has been found to have a bidirectional neuron-endocrine-immune system regulating effect, and antagonize systemic inflammatory response without side effects. We have demonstrated that electroacupuncture (EA) at ST36 had a significantly positive effect on hemorrhagic shock in rats with delayed fluid resuscitation, however, its mechanism remains unknown. We have recently furthermore proved that EA alleviated intestinal barrier insult and system inflammation in a rat ischemia model through activating the cholinergic anti-inflammatory pathway[17]. Therefore, we investigated whether EA at ST36 protected intestinal barrier function, thus preventing remote organ injury after prolonged hemorrhagic shock in rats with delayed fluid replacement (DFR) through activating the cholinergic anti-inflammatory-dependent mechanism.

MATERIALS AND METHODS

Animals

Male Sprague-Dawley rats (8-10 wk, 240-260 g) were purchased from Experimental Animal Center of Military Medical Sciences of the Chinese PLA. Rats were acclimatized for a while in mesh cages in a temperature-controlled room with a 12-h light-dark cycle in the animal quarter of our laboratory and fasted overnight, but allowed free access to water until 4 h before surgery. The research protocols were approved by the Committee of Scientific Research of the First Hospital Affiliated to General Hospital of PLA, China. The experiment was conducted in compliance with the Guide for Care and Use of Laboratory Animals of National Research Council, China.
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**Surgical procedures**

Rats were anesthetized and instrumented with 3% isoflurane inhalation (Yeean Technology Limited, Beijing, China). Ketamine 10 mg/kg was hypodermically injected for local anesthesia. Isoflurane (0.7%) was used to maintain anesthesia during the experiments. Animals were allowed to breathe spontaneously under a nose cone scavenging system using a veterinary anesthesia delivery system (Kent Scientific TOPO, Torrington, CT, United States). With aseptic technique, polyethylene (PE50) catheters were placed in the right carotid artery for continuous artery blood pressure monitoring, in the left femoral artery for blood withdrawal, and in the right femoral vein for fluid infusion. A 2-cm upper-midline laparotomy incision was made to identify gastroesophageal junction and expose the dorsal and ventral vagus nerve on the distal esophagus with a Phenix XLT165-LB stereomicroscope (Phenix Optical Instrument Group Company, Jiangxi Province, China). Rectal temperature was maintained at 37 °C with a heating pad and a heating lamp.

**Hemorrhagic shock protocol**

Each animal’s estimated blood volume was calculated using the formula \[ V = BW \times 0.06 \times 0.77 \text{ (mL/kg)} \]. Total blood volume (TBV) (mL) = body weight (g) × 0.06 (mL/g) × 0.77. Hemorrhagic shock was induced by withdrawing 45% of the calculated TBV within 20 min (30% was withdrawn over the first 3 min and suspended for 7 min, and the rest 15% was withdrawn over another 10 min), using an infusion or a withdrawal pump (Kelifeng Apparatus, Beijing, China). The completion of the hemorrhagic shock model concluded the preparation phase, and the time was metered 0. Mean arterial pressure (MAP) was monitored using a PICCO-PLUS cardio-pulmonary volume monitor (PULSION, Feldkirchen, Germany) after exsanguinations were initiated, and recorded 0.5 h before bleeding, immediately upon completion of the hemorrhagic shock (0), 3 h and 12 h after blood loss. The early survival rate was recorded at 12 h after bleeding. Fluid resuscitation was initiated 3 h after exsanguinations.

**Animal grouping and treatment**

All the animals underwent the same surgical procedure and hemorrhagic shock protocol, and the experimental rats were randomly assigned to six groups: (1) EAN group: Rats underwent EA at non-channel acupoints located 0.5 cm lateral and distal from ST36 points \[^{[19]}\], followed by DFR similar to EA group; EA parameters and delayed rehydration were the same as the EA group; (2) EA group: Animals underwent EA at ST36 points, which were located at posterior and lateral side of the knee joint, 5 mm below capitulum fibulae \[^{[10]}\], immediately after the blood loss followed by DFR 3 h after hemorrhage. EA at ST36 with an electro-acupuncture apparatus (HANS, LH202H) was performed as described before \[^{[17]}\]. Briefly, both hind limbs were shaved and the skin was disinfected. ST36 acupuncture point was punctured with a depth of 7 mm, and then the needle was connected with an electro-acupuncture apparatus. The electric current with the intensity of 2 mA and 2-100Hz was continued for 1.5 h immediately after hemorrhage. Three hours after blood loss, rats were given a femoral vein infusion with Ringer lactate (2 times the amount of blood loss); (3) VGX/EA group: Animals underwent vagotomy of the dorsal and ventral vagus nerve on the distal esophagus prior to EA at ST36 immediately after blood loss followed by DFR; EA parameters and delayed rehydration were the same as the EA group; (4) VGX/EAN group: Animals underwent vagotomy similar to VGX/EA group before EA at non-channel acupoints similar to EAN group immediately after blood loss followed by DFR. EA parameters and delayed rehydration were the same as the EA group; (5) α-BGT/EA group: α-bungarotoxin (α-BGT 1 µg/kg, an antagonist of α7 subunit of cholinergic nicotinic receptor, which inhibits the α7 subunit of acetylcholine receptors by blocking a pivotal communication pathway between the efferent vagus and intestinal immune cell \[^{[20,21]}\]) was injected intraperitoneally prior to hemorrhage and followed by EA at ST36 and DFR similar to EA group. EA parameters and delayed rehydration were the same as the EA group; and (6) α-BGT/EAN group: α-BGT was injected intraperitoneally prior to hemorrhage and followed by EA at non-channel acupoints and DFR similar to EAN group.

Each group was then randomly divided into two subgroups: one subgroup (n = 12) for investigation of survival rate and MAP, the other one (n = 18) for cytokine levels, organ parameters, gut injury score, ZO1 detection and intestinal permeability to FITC-dextran test; blood and intestine for cytokine levels were harvested at 0, 3 and 12 h after blood loss (3-5 animals per group); blood for organ parameters test, and intestine for gut injury score, ZO1 detection and intestinal permeability to FITC-dextran test were harvested at 12 h after blood loss (3-5 animals per group).

The scheme for whole experiment is as follows in Figure 1.

**Samples of blood and intestine**

Rats were anesthetized with 3% isoflurane inhalation. Blood was drawn through left femoral artery at 0, 3 and 12 h after blood loss, and then the animals were sacrificed for distal small intestine harvest. Plasma was obtained by centrifuging the blood at 10000 × g for 10 min at 4 °C. Organ functions were assessed by measuring blood amionotransferase (ALT), MB isoenzyme of creatine kinase (CK-MB) and creatinine (Cr) using a Cobas 6000 automatic biochemical analyzer (Roche Diagnostics, Basel, Switzerland). Segments of distal small intestine were harvested and immediately homogenized on ice with 1 mL denaturing lysis buffer or non-denaturing lysis buffer for Western blotting or ELISA. The homogenate was then centrifuged at 10000 × g for 10 min at 4 °C. Aliquots of the supernatants of plasma and tissue were stored at -80 °C until use. Segments of intestine were also harvested and fixed in 4% paraformaldehyde for histologic
evaluation and immunofluorescence.

**Detection of TNF-α, IL-6 and IL-10 levels in plasma and intestine**

TNF-α, IL-6 and IL-10 levels in the plasma and intestine were assessed using commercially available ELISA kit according to the protocol provided by the manufacturer (Nanjing Jiancheng Corp., Nanjing, China). Supernatants were transferred into fresh tubes for the evaluation. Briefly, after adding 50 μL assay buffer, 50 μL samples or standard concentration for TNF-α, IL-6 or IL-10 were incubated with 50 μL diluted Biotin-Conjugate for 2 h at room temperature. After 3 washes, the plates were incubated with Streptavidin-HRP for 1 h at room temperature. After 3 washes, 3,3',5,5'-Tetramethylbenzidine (TMB) substrate solution was added to the plates for 15 min and the reaction was stopped with stop solution. The absorbance rate was read at 450 nm. The concentrations of the samples were calculated according to the standard curve. The plasma TNF-α, IL-6 and IL-10 levels were expressed as pg/mL. Intestine TNF-α, IL-6 and IL-10 levels were expressed as picograms per milligram of protein.

**Histopathologic score**

The paraformaldehyde-fixed intestine was embedded in paraffin, and cut into 2-μm sections. Hematoxylin and eosin staining of the intestine was performed by the Pathology Department of the First Hospital Affiliated to the People’s Liberation Army General Hospital. Then the sections were viewed under a light microscope and evaluated by a pathologist blinded to the experimental groups. The injury to the intestinal mucosa was scored using the grading system as previously described.[20]

**Intestinal epithelial permeability**

An in vivo intestinal permeability assay was performed to assess gut barrier function as described by Kao et al.[23]. Briefly, 30 min before sacrifice, animals were anesthetized with inhaled isoflurane. A midline laparotomy incision was made and a 10-cm segment of distal ileum was isolated between silk ties. A solution of 1.0 mL phosphate-buffered saline (PBS, pH 7.2) containing 25 mg 4-kDa fluorescein isothiocyanate (FITC)-dextran (Sigma-Aldrich, St. Louis, MO, United States) was injected into the lumen of the isolated segment of intestine. The bowel was returned to the abdominal cavity and the abdomen was closed. Animals were maintained lightly under general anesthesia for 30 min, and systemic blood was drawn by left femoral artery puncture and placed in heparinized Eppendorf tubes on ice. Plasma was obtained by centrifuging the blood at 10000 × g for 10 min at -4 °C. Plasma fluorescence was measured by a fluorescence spectrophotometer (Synergy2; BioTek Multi-Detection Microplate reader, United States) and compared with a standard curve of known concentrations of FITC-dextran diluted in rat plasma.

**Immunofluorescence**

After deparaffinization, the intestine sections were rehydrated and incubated in citrate buffer (Zhongshan Jinqiao Biotechnology Co., Ltd., Beijing, China) for heat-induced antigen retrieval. After three washes with PBS, sections were incubated with 3% bovine serum albumin (BSA) (Zhongshan Jinqiao Biotechnology Co., Ltd., Beijing, China) for 30 min to block nonspecific binding sites. The sections were then incubated in the ZO-1 antibody (1:100; Life Technologies, Gaithersburg, MD, United States) at 4 °C overnight. The following day, after washing with PBS three times, they were treated with Alexa Fluor 488 secondary goat anti-rabbit antibody in 1% BSA for 1 h at room temperature. Prolong Fade (Antifade Mounting Medium, Beyotime Institute of Biotechnology, Beijing, China) was added on placement of cover slips. Images were viewed using the Olympus fluorescence microscope (BX51-DP71) with exposure-matched settings.

**ZO-1 expression**

The harvested gut tissues were placed in 1 mL lysis buffer (50 mmol/L Tris-HCl, pH 7.4; 150 mmol/L NaCl; 1% NP-40; 0.1% sodium dodecyl sulphate), then homogenized and centrifuged at 12000 × g for 10 min. Following centrifugation, the supernatant was collected and analyzed for protein concentration. Protein concentrations were determined using a protein assay kit (Applygen Technologies Inc, Beijing, China). Total protein (100 μg) was loaded onto a sodium dodecyl sulfate-polyacrylamide (SDS-PAGE) gel and run at 120 volts for 2 h. After electrophoresis, the protein was transferred to a polyvinylidene difluoride membrane (PVDF; Applygen Technologies Inc, Beijing, China) and blocked for 2 h in TBST (50 mmol/L Tris; 150 mmol/L NaCl; 0.05% Tween 20) containing 5% milk (Applygen Technologies Inc, Beijing, China). The membrane was then incubated with the primary antibodies against glyceraldehyde 3-phosphate dehydrogenase (GAPDH) (1:5000; Zhongshan Jinqiao Biotechnology Co., Ltd., Beijing, China), and ZO-1 (1:500; Life Technologies, Gaithersburg, MD, United States) at 4 °C overnight. After 3 washes in TBST, the membrane was incubated with corresponding secondary antibodies conjugated to horseradish peroxidase at room temperature for 30 min and chemiluminescence was detected using SuperECL Plus (Applygen Technologies Inc, Beijing, China). Films were developed using a standard photographic procedure. Quantitative analysis of detected bands was carried out by densitometer scanning (ImageJ).

**Statistical analysis**

Data were analyzed using a commercial statistical soft-
ware package (SPSS statistics 17.0). Continuous variables were expressed as mean ± SEM. Statistical significance of differences between groups was determined using one way analysis of variance (ANOVA) followed by Dunnett’s test and SNK-q for multiple comparisons. If some variables were abnormally distributed, the Kruskal-Wallis H test was used. Significance was declared for *P* values < 0.05.

### RESULTS

**Effect of EA ST36 on plasma and intestinal cytokine levels**

Figure 2 illustrates the effect of EA ST36 on TNF-α, IL-6 and IL-10 in plasma and intestine after blood loss. Blood samples and intestine were obtained at 0, 3 and 12 h after blood loss. Data are expressed as mean ± SD (3-5 animals per group at each time point). *P* < 0.05 vs EAN group; *P* < 0.05 vs 0 h in the same group. EA: Electroacupuncture; VGX: Vagotomy; α-BGT: α-bungarotoxin; TNF: Tumor necrosis factor; IL: Interleukin.

**EA ST36 decreased intestinal injury**

Histological evaluation of intestinal injury was performed at 12 h after blood loss. Sections of distal ileum from ani-

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| Figure 2 | Tumor necrosis factor-α, interleukin-6 and interleukin-10 levels in plasma and intestine at 0, 3 and 12 h after blood loss. Blood samples and intestine were obtained at 0, 3 and 12 h after blood loss. Data are expressed as mean ± SD (3-5 animals per group at each time point). *P* < 0.05 vs EAN group; *P* < 0.05 vs 0 h in the same group. EA: Electroacupuncture; VGX: Vagotomy; α-BGT: α-bungarotoxin; TNF: Tumor necrosis factor; IL: Interleukin. |
mals in EAN group showed villous tip necrosis, blunting, and sloughing of villi (Figure 3A). EA at ST36, which was applied immediately after blood loss, significantly attenuated the mucosal damage (Figure 3B). In contrast, when abdominal vagotomy or intraperitoneal injection of α-BGT was performed and as such the intact neur-enteric axis was interrupted, EA at both ST36 and non-channel acupoints failed to prevent the histologic changes induced by hemorrhagic shock in the gut (Figure 3C-F). These data demonstrated that an intact vagus nerve is required for the biological effect of EA at ST36 in protecting against gut injury.

Gut injury scores were measured in all 6 groups (Figure 4). Animals in EAN group had an average injury score that was significantly higher than in EA group (3.78 ± 0.59 vs 1.85 ± 0.33, \( P < 0.05 \)). Abdominal vagotomy or intraperitoneal injection of α-BGT before EA at ST36 eliminated the protective effects of EA at ST36 and resulted in similar scores as animals in VGX/EAN, α-BGT/EAN and EAN groups (3.9 ± 0.53, 4.1 ± 0.29 vs 3.78 ± 0.59; \( P > 0.05 \), respectively).

EA at ST36 lowered intestinal permeability

The intestinal permeability was evaluated in an in vivo assay using FITC-Dextran (Figure 5). Animals in EA group had a significantly lower level of plasma FITC-Dextran when compared with EAN group (856.95 ± 90.65 ng/mL vs 2305.62 ± 278.32 ng/mL, \( P < 0.05 \)). However, when abdominal vagotomy or α-BGT injection was performed before EA at ST36, the intestinal permeability...
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was indistinguishable from animals in EAN group, and animals in VGX/EAN or α-BGT/EAN group also showed no protection in reducing intestinal permeability compared with animals in EAN group (2249.87 ± 294.17 ng/mL and 2400.15 ± 203.15 ng/mL vs 2305.62 ± 278.32 ng/mL).

EA ST36 prevented loss and redistribution of ZO-1

The tight junction protein, ZO-1, undergoes protein expression alterations in response to hemorrhagic shock. Exposure-matched fluorescent intensity correlated to the amount of ZO-1 protein expression after immunostaining (Figure 6). After blood loss, animals in EAN group showed a loss and redistribution in ZO-1 expression evidenced by a low fluorescent intensity at the cell periphery (Figure 6A). Animals in EA group (Figure 6B) showed preservation of the robust structure of ZO-1 staining, whereas after vagotomy or injection of α-bungarotoxin, it eliminated such protection. All images are taken at × 400 magnification with black bar = 5 μm. (3-5 animals per group at 12 h after blood loss, size bar = 2 μm).

These results were confirmed by Western blotting for the ZO-1 protein in intestinal tissue lysates (Figure 7). When compared with the average relative band density of

Figure 6  Intestinal ZO-1 immunofluorescent staining at 12 h after blood loss. Animals in EAN group showed a low fluorescent intensity at the cell periphery after hemorrhagic shock, and electroacupuncture (EA) at ST36 showed preservation of the robust structure of ZO-1 staining, whereas after vagotomy or injection of α-bungarotoxin, it eliminated such protection. All images are taken at × 400 magnification with black bar = 5 μm. (3-5 animals per group at 12 h after blood loss, size bar = 2 μm).

Figure 7  Intestinal ZO-1 protein expression at 12 h after blood loss. Intestinal extracts were obtained from animals at 12 h after blood loss for measurement of ZO-1 protein expression using Western blotting. Representative Western blotting for the ZO-1 protein is shown with its corresponding glyceraldehyde 3-phosphate dehydrogenase loading control to demonstrate equal protein load in all lanes. Electroacupuncture at ST36 resulted in preservation of protein expression. Significant reduction in ZO-1 expression was seen in all the other groups. *P < 0.05 vs EAN group, (3-5 animals per group at 12h after blood loss). EA: Electroacupuncture; VGX: Vagotomy; α-BGT: α-bungarotoxin; GAPDH: Glyceraldehyde-3-phosphate dehydrogenase.
Table 1  Alanine aminotransferase, MB isoenzyme of creatine kinase and creatinine of rats of all groups after hemorrhagic shock with delayed fluid replacement

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<th>EA</th>
<th>VGX/EA</th>
<th>VGX/EAN</th>
<th>α-BGT/EA</th>
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<sup>a</sup>P < 0.05 vs EAN group; <sup>b</sup>P < 0.05 vs 0 h in the same group. (3-5 animals per group at each time point after blood loss). ALT: Alanine aminotransferase; CK-MB: MB isoenzyme of creatine kinase; Cr: Creatinine; EA: Electroacupuncture; VGX: Vagotomy; α-BGT: α-bungarotoxin.

Figure 8  Effect of electroacupuncture ST36 on mean arterial pressure in rats after hemorrhagic shock with delayed fluid replacement. *P < 0.05 vs EAN group; *P < 0.05 vs 0 h in the same group. (3-5 animals per group at 12 h after blood loss). EA: Electroacupuncture; VGX: Vagotomy; α-BGT: α-bungarotoxin.

animals in EAN group, animals treated with EA at ST36 had significantly higher ZO-1 expression (P < 0.05). In contrast, intestinal ZO-1 protein levels were significantly decreased in animals receiving abdominal vagotomy or α-BGT injection before EA at ST36. ZO-1 protein levels in VGX/EAN or α-BGT/EAN group receiving EA at non-channel acupoints after vagotomy or α-BGT injection were not significantly different from that in EAN group.

**EA ST36 lowered plasma ALT, CK-MB and Cr**

There was a significant decrease in ALT, CK-MB and Cr in the EA group compared with EAN group (P < 0.05; Table 1). However, there was no significant difference of ALT, CK-MB, BUN and Cr among the EAN, VGX/EA, VGX/EAN, α-BGT/EA and α-BGT/EAN groups.

**EA at ST36 improved the blood pressure and raised the survival rate**

MAP in each group was significantly lowered after blood loss. MAP at time 0 was only 35%-45% of that before blood loss; after 3 h, MAP in each group increased to different degrees. At 3 h and 12 h after hemorrhage, the EA group displayed higher MAP levels than the EAN group (P < 0.05) (Figure 8) while the VGX/EA, VGX/EAN, α-BGT/EA and α-BGT/EAN groups had low MAP levels similar to the EAN group.

As shown in our previous study, at 12 h after blood loss, the survival rate of the EA group was significantly higher than that of the other groups (P < 0.05). Ten (83.3%) of 12 rats in the EA group were alive after blood loss. In contrast, only 5 (41.7%) of 12 rats in the EAN group survived after being subjected to hemorrhagic shock treated with EA at non-channel acupoints and DFR. When rats subjected to hemorrhagic shock were treated with EA at ST36 after abdominal vagotomy or α-BGT injection, 6 (50%) of 12 rats in VGX/EA group, 5 (41.7%) in α-BGT/EA group, 5 (41.7%) in VGX/EAN group and 5 (41.7%) in α-BGT/EAN group survived 12 h after blood loss.

**DISCUSSION**

The most important novel findings from this study...
are that when delayed resuscitation is inevitable during emergency situations such as hemorrhagic shock in war without sufficient fluids, EA at ST36 can be performed and it can successfully attenuate systemic inflammation, decrease gut injury and permeability and improve blood pressure and outcomes, which is consistent with preserved intestinal barrier function after hemorrhage and delayed fluid resuscitation. Based on our previous studies from this lab\cite{2,24}, these results have provided further evidence for a role of acupuncture in an emergency treatment after shock and trauma.

It is regarded that the intestinal tract is one of the earlier organs involved in ischemia-reperfusion injury after hemorrhagic shock. In order to maintain the blood supply of the vital organs during hemorrhagic shock, the intestinal blood flow sharply reduced, and dysfunction of the intestinal mucosal barrier occurs. The ischemia of the small intestine leads rapidly to an impairment of mucosa barrier function, and thus the earliest restitution of the mesenteric blood flow is essential. However, in a very harsh environment with a lack of decent medical support, such as accident and war, immediate resuscitation is sometimes unavailable, the delayed fluid resuscitation occurs, threatening the life of those with extensive injury or hemorrhagic shock. It has also been recognized that a delay in such replenishment could sometimes be fatal due to complications subsequent to delayed resuscitation of hypovolemic shock. One strategy for reducing such a hazard is to try to supplement liquid with sufficient electrolytes by mouth until intravenous infusion fluids are available\cite{25}. In recent years, we have been working on the alternative methods to treat the complications caused by delayed fluid resuscitation. We have found that oral resuscitation is an effective way to partly replace immediate resuscitation when the intravenous infusion is not available\cite{26}. More recently, we have demonstrated that EA at ST36 can significantly improve the survival rate and blood pressure after fatal hemorrhagic shock in rats\cite{27,28}.

The gut barrier has been found to be seriously damaged at the early phase of hemorrhagic shock\cite{29}. The protective effect of traditional Chinese medicine against intestinal and gastric mucosal injury after hemorrhagic shock in rats has been investigated\cite{27,28}. Increasing evidences suggest that the effect of EA at ST36 for gastrointestinal disorders may involve vagal reflex. The dorsal vagal complex (DVC) consists of the nucleus of the solitary tract (NTS), which receives primary visceral afferent information, and the dorsal motor nucleus of the vagus (DMV), which contains the efferent vagal neurons innervating visceral organs. Therefore, DVC plays an important role in regulating visceral functions. A previous study has demonstrated that there is a commonality of central nervous system (CNS) cell groups in brain controlling ST36 point, including DMV and NTS\cite{28}. EA at ST36 generated an increased c-Fos expression in the neurons of DMV\cite{29} and promoted the gastric myoelectric activity, which was regulated by the vagus, and substance P (SP) which is widely present in DVC and involved in the excitatory effects\cite{30}. It has been proved that EA at ST36 can increase the efferent activity of the vagal nerve\cite{31}. Taken together, these data suggest that EA at ST36 is relevant to vagus nerve and it has beneficial effects in intestinal barrier function. In this set of experiments, we stimulated ST36 points after hemorrhagic shock, established its efficacy, and compared its effects with EA at ST36 or non-channel acupoints after abdominal vagotomy or α-BGT injection. We demonstrated that EA at ST36 is effective in preventing intestinal barrier breakdown after hemorrhagic shock with delayed resuscitation. We also showed, as a proof of concept, that its biological effect is dependent on an intact vagus nerve.

Increasing evidence points to extensive cross-talk between intestinal barrier breakdown and cytokine overproduction. The gut has been shown to be a source of inflammatory cytokine with capability of priming neutrophils after ischemia injury or hemorrhagic shock\cite{32,33}. These cytokines originating from the gut may then exacerbate the systemic inflammatory response and potentially lead to a further damage to gut permeability\cite{34,35}. It has also been demonstrated that IL-6 is essential for the development of gut barrier dysfunction after hemorrhagic shock\cite{35}. IL-10 has also been proved to play a pivotal role in regulating proinflammatory cytokine release following trauma-hemorrhage\cite{36}. In our previous studies, we demonstrated that EA at ST36 can alleviate intestinal pro-inflammatory factors, tissue edema and insult of intestinal mucosa\cite{37}, significantly protect against tumor necrosis factor-α induced-multiple organ dysfunction in rats with sepsis\cite{38}, and promote gastric emptying in rats with a 40% blood volume loss\cite{39}. EA at ST36 has been shown to have a regulatory effect on TNF-α level in rats\cite{40}. This study also demonstrated that acupuncture at ST36 significantly attenuated the expression of pro-inflammatory cytokine TNF-α and IL-6 levels in plasma and intestinal homogenate, but failed to suppress the anti-inflammatory cytokine IL-10 levels in plasma and intestine. These data suggests the anti-inflammatory potential of the use of ST36 acupuncture against hemorrhagic shock. Non-channel acupoints acupuncturing, vagotomy or α-BGT injection before EA at ST36 showed no effects in reducing the production of TNF-α and IL-6, which suggested that anti-inflammatory effect of EA at ST36 may be exerted through the vagal nerves, and their integrity is essential.

In our previous study, we demonstrated that EA at ST36 can effectively protect organ function\cite{24}, improve the early survival rate, increase the intestinal tissue diamine oxidase activity and alleviate intestinal ischemia in rats with DFR after hemorrhagic shock\cite{25}, but its mechanism is unknown. This study demonstrated that this protective effect of EA at ST36 may be related to its protection of intestinal barrier function. In this study, EA at ST36 effectively prevented histologic injury of the gut mucosa, reduced permeability of the distal ileum to 4-kDa FITC-dextran and maintained intestinal tight junction protein expression and function. We also proved that this biologi-
Electroacupuncture improves gut barrier dysfunction in hemorrhagic shock rats

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A delayed resuscitation are of great significance in controlling hemorrhagic shock and subsequent organ dysfunction when hemorrhagic shock prolonged due to systemic inflammatory response syndrome (SIRS) and the multiple casualties occurring in austere circumstances such as in the battlefield or site of an unexpected accident or a disaster. Acupuncture as one of the therapeutic maneuvers in traditional Chinese medicine (TCM) has been applied in clinics for thousands of years, and it has been found to have a bidirectional neuron-endocrine-immune system regulating effect, and antagonize the systemic inflammatory response without side effects. The authors demonstrated that electroacupuncture (EA) at Zusanli had a significant positive therapeutic effect in hemorrhagic shock rats with delayed fluid resuscitation, however, its mechanism remains unknown. The authors had also proved that EA can alleviate intestinal barrier insult and systemic inflammation in a rat ischemia model through activating the cholinergic anti-inflammatory pathway.

Innovations and breakthroughs

The most important novel findings from this study are that when delayed resuscitation is inevitable during emergency situations such as hemorrhagic shock occurring in the battlefield or site of an unexpected accident or a disaster without sufficient fluids, EA at ST36 can be performed and it can successfully attenuate systemic inflammation, decrease gut injury and permeability and improve blood pressure and outcomes, which is consistent with preserved intestinal barrier function after hemorrhage and delayed fluid resuscitation.

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Comments

Background

Fluid resuscitation for hemorrhagicburn shock is often challenging when mass casualties occurring in austere circumstances such as in the battlefield or site of an unexpected accident or a disaster, where intravenous fluid resuscitation is difficult or even impossible. Subsequent to delayed resuscitation of hypovolemic shock, a high mortality and an increase in incidence of serious complications such as systemic inflammatory response syndrome (SIRS) and the multiple organ dysfunction syndrome (MODS) may thus befall to these victims. Thus, interventions such as drug or acupuncture to prevent excessive inflammation and subsequent organ dysfunction when hemorrhagic shock prolonged due to delayed resuscitation are of great significance in controlling hemorrhagic shock without sufficient fluid infusion, especially during evacuation and transportation.

Research frontiers

The current treatment for hypovolemic shock focuses on maintaining sufficient tissue perfusion and vital organ function with early and adequate fluid replenishment. It is of great importance to seek clinically alternative therapies when delayed fluid replenishment is inevitable. Activation of the cholinergic anti-inflammatory pathway by vagus nerve stimulation has been demonstrated to inhibit inflammatory response, prolong survival and protect against the development of hypotension during lethal hemorrhagic shock. Gut, especially the intestinal barrier function, plays a key role in the development of intestinal and systemic inflammatory response and protection of organ function following hemorrhagic shock. Researchers have demonstrated an expanded role for vagus nerve stimulation and the cholinergic anti-inflammatory mechanism that provides a protective effect on the gut against epithelial barrier dysfunction and alleviates inflammatory injury in intestine and remote organs. However, due to complicated manipulation and untoward side effects, including serious tissue injury, it is still difficult to apply electrical stimulation to the vagus nerve in clinical practice. Therefore, a more clinically desirable alternative therapy is needed during the resuscitative phase of trauma care. Acupuncture as one of the therapeutic maneuvers in traditional Chinese medicine (TCM) has been applied in clinics for thousands of years, and it has been found to have a bidirectional neuron-endocrine-immune system regulating effect, and antagonize the systemic inflammatory response without side effects. The authors demonstrated that electroacupuncture (EA) at Zusanli had a significant positive therapeutic effect in hemorrhagic shock rats with delayed fluid resuscitation, however, its mechanism remains unknown. The authors had also proved that EA can alleviate intestinal barrier insult and system inflammation in a rat ischemia model through activating the cholinergic anti-inflammatory pathway.

Applications

The study results provide evidences for acupuncture as an emergency intervention of anti-shock and protection of gut barrier function after severe hemorrhage and trauma.

Terminology

Cholinergic anti-inflammatory pathway: The cholinergic anti-inflammatory pathway is a neural mechanism that inhibits the expression of cytokine through the interaction of the principle vagus nerve neurotransmitter, acetylcholine, and the α7nAChR subunit located on vagus nerve expressing cells by stimulating the vagus nerve by either electrical or pharmacological methods. EA: EA is a modification of conventional acupuncture that stimulates acupoints with electrical current instead of manual manipulations and appears to have more consistently reproducible results in both clinical and research settings.

Peer review

The study is somewhat interesting, well designed and results seem to be very congruent among the different aspects of systemic inflammation and intestinal barrier function investigated.

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